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Chemical Compositions and Physical Characteristics of Volatile Extracts of Leaves of *Psidium Guajava* Linn and *Lantana Camara* Linn of Benin

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ABSTRACT

P. guajava Linn. *L. camara* Linn. are two aromatic plants that produce essential oils and major medicinal utilities in Benin. Essential oils obtained by hydrodistillation of *P. guajava* and *L. camara* leaves collected in Benin were analyzed by gas chromatography connected to a flame ionization detector (GC/FID) and gas chromatography coupled with the mass spectrometry (GC/MS). The major compounds (> 10%) identified in the essential oils of *P. guajava* are limonene (10.9-20.7%), β -bisabolol (14.9-20.2%), epi- β -bisabolol (11.7-18.9%), (2E,6E)-farnesol (10.0%), β -bisabolene (10.0%), 1,8-cineole (23.4%), sabinene (15.9-21.5%), β -caryophyllene (10.9-19.3%), (Z)- β -ocimene (11.2-13.8%) were the major constituents (>10%) having marked the essential oils of *L. camara*. The physical characteristics determined have varied, at the same time, according to the plant species and within the samples forming a homogeneous population botanically.

Keywords: *Psidium guajava*, *Lantana camara*, epi- β -bisabolol, 1,8-cineole, density, refractive index, rotary power, Benin.

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INTRODUCTION

The *P. guajava* Linn. is a small tree (5 to 8m high), hard wood and tortuous, a native of Central America [1-4], but knows a good development in the tropical regions of Africa [5]. In the nutritional plan, the fruit of *P. guajava*, similar in form to that of aspherical berry is very edible [6]. It is an excellent source of vitamins essential for growth and proper functioning of the human body (especially vitamins C and A) [7]. To Cambridge and in China, the leaves are used to reduce fever, to restore tonic psychiatrist and to relieve the diabetes [8,9]. Its aerial parts and roots, rich in tannins have astringent and anti-diarrheal properties [10]. Moreover, Petard reported in 1986 that Tahitiens of the French Polynesia considered, 150 years ago, the *P. guajava* as a Polynesian medicinal plant [11]. In traditional medicine in Benin, *P. guajava* helps to treat dysentery, diarrhea and jaundice [5].

Several scientific studies relating to the determination of the chemical composition of *P. Guajava* and it has been reported for various compounds characteristic of the type of essential oil studied. In 1991, Xiao-duo *et al.* have shown the existence, an important proportion, of 1,8-cineole (18.9%) and α -pinene (37.8%) in the essential oil of *P. guajava* leaves acclimated in China [9]. These same main compounds (monoterpenoids) were identified in the volatile extracts of *P. guajava* leafy-stems of Brazil (1,8-cineole: 21.4%, α -pinene: 23.9%) [3] and Taiwan (1,8-cineole: 12.4%, α -pinene: 14.7%) [4]. Sagrero-Nieves in Mexico (1994) [12], Jorge in Cuba [1], Ogunwandé in Nigeria (2003) [2], Chen in Taiwan (2007) [4] and Soares in Brazil (2007) [7] have published some chemical compositions marked by the presence of other major constituent such as α -selinene, β -caryophyllene, δ -selinene, (E)-nerolidol, selin-11-en-4- α -ol, limonene, (E)-2-hexenal and *cis*-hex-3-en-1-ylacetate.

Benin is a tropical country with lush vegetation, rich in aromatic plant species varied. *L. camara*, part of this vegetation is aromatic and essential oil product. It is of the verbenaceae family and it grows in all the tropical and subtropical regions of America, Africa and Asia. This tree carries triangular leaves at the top and regularly toothed on the edges. The leaves have enough virtue recognized in Benin traditional medicine. Indeed, Adjanohoun reported in 1989 [5] that the *L. camara* stem-leaves, combined with those of *Caesalpinia pulcherrima* relieve schizophrenia. He also indicated that the decoction of *L. camara* leaves and roots, associated with the barks of *Mangifera indica* and *Anacardium occidentale*, then the *Olexis ubscorpioidea* roots and *Xylopi aethiopica* fruits, is used orally against diabetes [5]. Essential oils extracted from the leaves, seeds and flowers of *L. camara* was the object of several works since a few years [13, 14]. Recently in 2004, Alitonou reported the presence in the leaves harvested to Abomey by important compounds such as β -caryophyllene (18.5%), sabinene (13.1%), α -humulene (10.0%), 1,8-cineole (9.0%) and δ -guaïene (5.0%) [15]. Later in 2005, Randrianalijaona has emerged major compounds of the essential oil of pink-purple flowers of *L. camara*, collected in Madagascar (sabinene (9.4-11.3%), 1,8-cineole (3.7-4.6%), linalool (4.8-6.1%), β -caryophyllene (11.3-13.6%), α -humulene (0.1-0.4%), β -bisabolene (1.7-2.3%), δ -cadinene (0.1-0.4%), α -curcumene (1.0-1.6%), caryophyllene oxide (1.2-0.7%) and davanone (22.6-25.9%) [16]. Of such compounds in different proportions, were also noticed by GC/MS in the essential oils from *L. camara* leaves of Bangladesh [17], Brazil [18], India [19].

This study was prompted because of the increased interest of the population in these plants in the traditional medicine and in post-harvest systems in Benin. Unfortunately, the ecosystems usually harboring these plants disappear gradually under the influence of the strong demographic pressure and due to the absence of conservation policy of the virtuous plants. This work objective was to analyze and study by GC/MS the chemical composition of essential oils from leaves of *P. guajava* and *L. camara* then identify similarities and differences between the values of the physical properties (density, refractive index and rotatory power) of these volatile extracts.

MATERIALS AND METHODS

EXPERIMENTAL

Plant material and distillation of the volatile constituents

L. camara and *P. guajava* leaves were collected respectively in 2006 and 2007 in several different locations from the south of Benin. They were identified and certified to the Abomey-Calavi University National Herbarium and stored in the laboratory between 18 and 20°C in the shade of the sunlight throughout the extraction period. The essential oils were extracted by hydrodistillation of the leaves (250-300 g) for three hours on Clevenger according to the method used in British Pharmacopoeia [20]. They were dried over anhydrous sodium sulfate and analyzed by GC/MS.

Physical properties

Density at 20°C

The density measurement was carried out using a micro-pycnometer and a precision balance.

Refractive index at 20 ° C

The refractive index was determined by means of the refractometer CARL ZEISS JENA 234678.

Rotatory power at 20 ° C

The measurement was made by CARL ZEISS polarimeter 128291.

Analysis of the volatile constituents

GC/MS: The essential oils were analysed on a Hewlett-Packard gas chromatograph Model 7890, coupled to a Hewlett-Packard MS model 5875, equipped with a DB5 MS column (30m X 0.25mm; 0.25µm), programming from 50°C (5 min) to 300°C at 5°C/min, 5 min hold. Helium as carrier gas (1.0 mL/min); injection in split mode (1:30) ; injector and detector temperature, 250 and

280°C respectively. The MS working in electron impact mode at 70 eV; electron multiplier, 2500V; ion source temperature, 180°C; mass spectra data were acquired in the scan mode in *m/z* range 33-450.

GC/FID: The essential oils were analysed on a Hewlett-Packard gas chromatograph Model 6890, equipped with a DB5 MS column (30m X 0.25mm; 0.25µm), programming from 50°C (5min) to 300°C at 5°C/min, 5min hold. Hydrogen as carrier gas (1.0 mL/min); injection in split mode (1:60); injector and detector temperature, 280 and 300°C respectively. The essential oil is diluted in hexane: 1/30.

The compounds assayed by GC in the different essential oils were identified by comparing their retention indices with those of reference compounds in the literature and confirmed by GC-MS by comparison of their mass spectra with those of reference substances [21, 22, 23].

RESULTS

The hydrodistillation made from the leaves of *P. guajava* and *L. camara* generated essential oils yield values between 0.018 and 0.82%.

Table 1 : Essential oil yield of leaves of *P. guajava* and *L. camara*

samples	<i>P. guajava</i>					<i>L. camara</i>		
	F ₁	F ₂	F ₃	F ₄	F ₅	F ₆	F ₇	F ₈
yield (%)	0.30±0.01	0.54±0.01	0.41±0.01	0.25±0.02	0.82±0.01	0.02±0.001	0.02±0.002	0.018±0.002
F ₁ = Adjarra (17-05-07), F ₂ = Banigbe (27-05-07), F ₃ = Hounsa (24-04-07), F ₄ = Misserete (19-08-07), F ₅ = Tchaada (26-08-07), F ₆ = Abomey-Calavi (07-06-06), F ₇ = Ouando (07-06-06), F ₈ = Seme (07-06-06)								

In the table 2 were grouped the experimental values of density, refractive index and rotatory power of *Psidium guajava* and *Lantana camara* leaves essential oils.

Table 2 : physical characteristics of *P. guajava* L. and *camara* leaves essential oils

samples	density (20°C)	refractive index(20°C)	rotatory power(20°C)
F ₁	0.961	1.4930	-12.7
F ₂	0.874	1.4945	-7.5
F ₃	0.732	1.4900	-12.3
F ₄	0.891	1.4901	-15.8
F ₅	0.926	1.4920	-10.1
F ₆	0.908	1.4735	5.2
F ₇	0.881	1.4715	5.3
F ₈	0.885	1.4705	5.3
F ₁ = Adjarra (17-05-07), F ₂ = Banigbe (27-05-07), F ₃ = Hounsa (24-04-07), F ₄ = Misserete (19-08-07), F ₅ = Tchaada (26-08-07), F ₆ = Abomey-Calavi (07-06-06), F ₇ = Ouando (07-06-06), F ₈ = Seme (07-06-06)			

The compounds identified by gas chromatography coupled with mass spectrometry and their Kovats indices are sorted by essential oils sample in the Table 3.

Table 3 : Chemical compositions of the essential oils of *P. guajava* and *L. camara* leaves

Components	R _{exp}	R _{lth}	(%)							
			<i>P. guajava</i>					<i>L. camara</i>		
			F ₁	F ₂	F ₃	F ₄	F ₅	F ₆	F ₇	F ₈
(3E)-hex-en-1-ol	854	844	-	-	-	-	0.6	-	-	-
tricyclene	929	921	-	-	0.9	1.7	0.3	0.4	0.4	-
α-thujene	932	924	-	-	-	-	-	-	2.9	4.0
α-pinene	935	932	-	-	-	-	-	2.3	-	-
camphene	946	946	-	-	-	-	-	1.2	-	1.6
α-fenchene	947	945	-	-	-	-	-	-	1.4	-
benzaldehyde	959	952	1.7	2.1	1.6	2.5	5.7	-	-	-
sabinene	973	969	-	-	-	-	-	15.9	18.4	21.5
β-pinene	979	974	-	-	-	-	-	1.9	2.3	3.4
6-methyl-5-hept-5-en-2-one	981	-	-	-	-	0.2	1.1	-	-	1.0
myrcene	991	988	0.3	-	-	0.2	-	1.8	1.9	2.1
p-mentha-1(7),8-diene	999	1003	-	-	-	0.3	-	2.5	2.3	2.6
α-terpinene	1015	1014	-	-	-	-	-	0.3	0.2	0.2
p-cymene	1021	1020	-	-	-	0.3	-	-	0.2	0.6
limonene	1028	1024	16.0	3.4	10.9	20.7	4.0	0.2	-	-
1,8-cineole	1029	1026	-	0.5	-	0.1	6.4	1.6	1.7	23.4
(Z)-β-ocimene	1037	1032	-	-	-	-	0.4	11.2	13.8	-
(E)-β-ocimene	1050	1044	-	-	-	-	0.3	1.2	1.4	1.0
γ-terpinene	1057	1054	-	-	-	-	-	0.6	0.5	0.8
cis-sabinene hydrate	1070	1065	-	-	-	-	-	0.9	1.1	0.6
terpinolene	1083	1086	-	0.3	-	-	-	0.4	0.5	0.4
linalool	1097	1095	-	-	-	-	0.3	-	-	-
trans-sabinene hydrate	1098	1098	-	0.3	-	-	-	0.9	0.6	0.8
cis-p-menth-2-en-1-ol	1122	1118	-	-	-	-	-	-	-	0.1
camphor	1146	1141	-	-	-	0.7	-	1.4	1.7	2.1
borneol	1171	1165	-	-	-	-	-	0.2	0.7	0.3
terpinen-4-ol	1177	1174	-	-	-	-	0.3	1.9	1.2	2.5
α-terpineol	1192	1186	-	-	-	-	0.9	1.1	1.1	1.1
methyl geranate	1316	1322	-	-	-	0.2	-	-	-	-
eugenol	1348	1356	-	-	-	-	-	1.3	-	-
α-copaene	1374	1374	2.4	2.7	2.3	3.9	1.0	0.4	0.3	0.4
β-elemene	1384	1389	-	-	-	-	-	0.6	0.5	0.3
sesquithujene	1398	1405	0.3	0.4	0.3	-	-	-	-	-
α-cis-bergamotene	1409	1411	0.9	-	1.2	-	0.5	-	-	-
β-caryophyllene	1419	1417	3.0	11.3	3.0	6.3	6.4	19.3	17.7	10.9
β-copaene	1428	1430	-	-	-	-	-	0.4	0.4	0.4
α-trans-bergamotene	1429	1432	0.5	0.7	0.5	0.3	0.7	-	-	-
(Z)-β-farnesene	1447	1440	0.4	0.5	0.4	-	0.5	-	-	-
α-humulene	1454	1452	0.6	1.6	0.6	1.2	1.0	6.5	5.9	4.4
β-santalene	1456	1457	0.6	0.5	0.7	-	0.6	-	-	-
sesquisabinene	1459	1459	-	-	-	-	-	0.2	-	-



allo-aromadenderene	1461	1460	-	-	-	-	-	-	0.2	-
α -acoradiene	1462	1464	0.9	1.6	1.8	-	1.1	-	-	-
β -acoradiene	1471	1469	-	2.5	1.3	-	2.5	-	-	0.1
γ -muurolene	1472	1478	-	-	-	0.5	-	-	0.2	0.9
α -curcumene	1474	1479	3.1	2.6	4.7	-	2.7	-	-	-
γ -curcumene	1478	1481	1.3	2.4	-	-	-	-	-	-
germacrene-D	1480	1484	-	-	-	-	1.2	0.2	1.8	-
β -selinene	1488	1489	0.5	1.7	0.5	3.1	-	-	-	-
α -zingiberene	1489	1493	-	0.6	-	-	-	-	-	-
α -selinene	1491	1498	0.5	0.5	-	2.8	0.7	-	-	3.1
bicyclogermacrene	1493	1500	-	-	-	-	-	1.4	6.8	-
(Z)- α -bisabolene	1494	1506	1.4	3.3	-	-	-	-	-	-
germacrene-A	1498	1508	-	-	1.4	-	-	6.0	0.2	0.1
γ -cadinene	1510	1513	0.9	0.7	1.3	-	-	-	-	0.4
β -curcumene	1512	1514	9.5	11.7	10.0	-	11.8	-	-	-
β -sesquiphellandrene	1520	1521	1.9	3.5	1.9	-	3.2	-	-	0.2
δ -cadinene	1522	1522	-	1.3	-	0.8	0.5	0.6	0.5	-
(E)- γ -bisabolene	1523	1529	1.9	2.6	1.9	-	2.8	-	-	-
α -cadinene	1536	1537	0.4	0.6	0.4	-	0.3	-	-	0.1
δ -cuprenene	1546	1542	-	-	-	-	-	0.9	0.5	0.2
germacrene-B	1555	1559	-	-	-	-	-	1.0	0.6	-
(E)-nerolidol	1556	1561	3.2	3.5	3.6	0.9	3.6	3.5	2.5	1.3
caryophyllene alcohol	1574	1570	-	-	-	0.4	-	0.6	-	-
sesquisabinene (E)-hydrate	1578	1577	-	-	-	-	-	0.6	0.3	0.6
caryophyllene oxide	1580	1582	0.9	1.9	1.1	2.7	1.2	0.8	0.6	-
β -copaen-4- α -ol	1584	1590	-	0.3	-	0.3	-	-	-	-
guaïol	1591	1600	-	0.9	-	0.4	-	-	-	-
epi-globulol	1606	-	0.6	1.7	0.6	0.4	0.5	-	-	-
humuleneepoxyde II	1607	-	-	0.6	-	1.2	-	0.8	0.6	0.5
1,10-di-epi-cubenol	1619	1618	-	1.5	-	0.3	1.9	-	-	-
1-epi-cubenol	1631	1627	1.9	-	4.6	3.7	-	2.1	-	1.3
α -acorenol	1632	1632	2.6	1.8	-	-	2.0	-	-	-
β -acorenol	1635	1636	1.3	1.3	1.3	2.4	-	-	-	-
epi- α -cadinol	1639	1638	1.2	2.0	1.4	-	2.3	0.4	0.2	-
epoxy-allo-alloaromadendrene	1633	1639	-	-	-	1.6	-	-	1.5	-
epi- α -muurolol	1641	1640	0.4	-	0.5	1.4	-	-	-	-
α -muurolol	1644	1644	-	1.1	1.3	2.3	1.0	-	-	-
cubenol	1653	1645	1.1	0.4	0.8	-	-	-	-	-
α -cadinol	1654	1652	-	1.4	1.3	-	1.0	-	-	-
neo-intermedeol	1658	1658	2.7	2.6	2.5	-	-	-	-	-
selin-11-en-4- α -ol	1660	1659	-	-	-	9.9	2.7	-	-	-
14-hydroxy-9-epi-(E)-caryophyllene	1668	1668	-	-	-	0.9	-	-	-	-
epi- β -bisabolol	1670	1670	-	-	20.2	-	15.8	-	-	-
β -bisabolol	1671	1674	18.9	11.7	-	-	-	-	-	-
nerolidyl acetate	1678	1676	1.1	0.8	1.2	-	1.4	1.3	1.0	-
epi- α -bisabolol	1680	1683	3.7	1.4	4.2	-	3.0	-	-	-
α -bisabolol	1683	1685	2.1	2.4	2.1	0.2	-	-	-	-
(2Z, 6Z)-farnesol	1709	1698	1.3	-	1.5	4.3	0.8	-	-	-
(2Z, 6E)-farnesol	1710	1714	0.7	0.5	0.6	10.0	-	-	-	-

(2E, 6E)-farnesol	1742		0.9	-	0.8	6.0	0.3	-	-	-
benzyl benzoate	1764	1759	-	0.4		0.5	-	-	-	-
<p>F₁= Adjarra (17-05-07), F₂= Banigbe (27-05-07), F₃= Hounsa (24-04-07), F₄= Misserete (19-08-07), F₅= Tchaada (26-08-07), F₆= Abomey-Calavi (07-06-06), F₇= Ouando (07-06-06), F₈= Seme (07-06-06), RI = Retention index, exp = experimental, th = theoric</p>										

DISCUSSION

The values of the yields of *P. guajava* essential oil recorded differ between them and are situated between 0.25% and 0.82%. The lowest yield of *P. guajava* leaves essential oils studied was 0.25%. It was approximately 2.5 times larger than that (0.1%) of *P. guajava* leaves harvested in French Polynesia [10]. The essential oil sample F₅ presented a proportion of essential oil (0.82%) remote from that (0.75%) leaves studied in Nigeria [2]. For cons, the essential oil yields (0.41%) of the sample F₃ close to that (0.4%) of *P. guajava* volatile extract studied in Brazil [3]. As for the *L. camara* leaves, the yields in essential oils of samples F₆, F₇, F₈ were lower than those generated by the samples of *P. guajava* essential oils. They were estimated, on average at 0.2%. According to the results of other studies reported in the literature on the genre *Lantana*, the essential oil yields of the aerial parts were between 0.01 and 0.4% [24, 25, 26].

In Table 2 were recorded the results of the determination of the physical properties (density, refractive index, rotatory power) of *P. guajava* and *L. camara* volatile extracts. Differences were observed in relation to various harvest sites on the one hand and on the other hand, the botanical species considered. The sample of essential oil of *P. guajava* F₁ appears as the most dense (0.961). The values of the rotatory power recorded were different from a botanical species each other. Consequently, the essential oil samples from the same botanical species (*P. guajava* or *L. camara*) exert the same action towards the polarized light. Concerning the refractive index, the values recorded were practically homogeneous by botanical species. The table 3 reports the compounds, with character fowl, identified in *P. guajava* and *L. camara* essential oils. At the level of the five samples of *P. guajava* essential oil, 40 to 51 compounds were highlighted representing 93.6% to 97.2% of the weight of the volatile extract studied. In this table, it appears that the essential oils from *P. guajava* leaves contain significant rates of hydrogenated and oxygenated sesquiterpenes (62.1-89.1%). These sesquiterpenic hydrocarbons (18.6-52.6%) were more represented than hydrogenated monoterpenes proportions more lower (3.4-23.3%). As for the oxygenated monoterpenes, their proportions in the essential oils studied were evaluated between 0.5% and 7.6%. The main products which form these samples of the essential oil were limonene (3.4-20.7%), β -bisabolol (14.9-20.2%), epi- β -bisabolol (11.7-18.9%), β -caryophyllene (6.1-10.9%), β -bisabolene (7.5-10%), (2E,6E)-farnesol (10.0%) and selin-11-en-4- α -ol (9.9%). It was also noted, with significant proportions, the presence of neryl (Z)-3-hexanoate (5.8%) in F₄ and α -cuprenene (7.2%) in F₅. The limonene and β -caryophyllene percentages of essential oil samples chromatographed were taken away well from those (respectively 42.1% and 21.3%) stemming from Ogunwande et al. (2003) work in Nigeria [2]. The samples F₁ and F₃ were devoid of 1,8-cineole, linalool and α -terpineol. Except 1,8-cineole (6.4%) found in F₅, linalool and α -terpineol occurred at less than 1% in the samples F₂, F₄, F₅. The F₅ proportion in 1,8-cineole was much lower than that (18.9%) noted in the volatile extract of *P.*

guajava leaves chromatographed in China [9] and more lower compared to the results published in BrasiliabydaSilvaet *al.*(2003) [3]. On the over hand, the results of the work published by various authors coming from Cuba in 2001, Nigeria in 2003, Taiwan in 2007 and Brasilia in 2007 did not mention the presence of 1,8-cineole in the group of odoriferous compounds of the essential oil of the leaves of *P.guajava* [1, 2, 4, 7]. Several esters have marked their presence in the essential oil of both aromatic species involved in this study. The neryl (Z)-3-hexanoate and isobornyl 5-hydroxy isobutyrate appeared with substantial proportions, respectively 5.8% and 2.5%. Other esters (acetate (Z)-nerolidyl, (2E,6E)-farnesyl acetate, benzyl benzoate, neryl benzoate, methylbutyl 2-methylbutanoate, isopentenyl isovalerate, 3Z-hexenyl butanoate, methylgeranate, 2-methylbutyl benzoate, neryl crotonate, geranyl 2-methylbutanoate) were indicated in traces (< 0.5%). It was important to note that the chemical compositions presented in this work highlighted the major compounds different from those identified in Brasilia (α -pinene: 23.9%; 1,8-cineole: 21.4%; β -bisabol: 9.2%) [3] and Mexico (α -selinene: 23.7%; β -caryophyllene: 18.8% and δ -selinene: 18.3%) [12]. According to the results in the Table 3, *L. camara* essential oils were marked by the presence of important proportions of hydrogenated and oxygenated monoterpenes (48.3-68.7%). The proportions of hydrogenated and oxygenated sesquiterpenes varied between 23.9 and 46.3%. The major compounds recorded independently of the origin of each of the samples of essential oils were 1,8-cineole (23.4%), sabinene (15.9-21.5%), (Z)- β -ocimene (11.2-13.8%), β -caryophyllene (10.9-19.3%), α -humulene (5.9-6.5%), bicyclogermacrene (6.8%), germacrene-A (5.8%), *cis*-prenyl limonene (4.3%) and α -thujene (4.0%). Unlike chemical profile samples F₆ and F₇ which were marked by traces, the rate in 1,8-cineole (23.4%) of fraction F₈ was very high. Let us say that the proportions of (Z)- β -ocimene (11.2% and 13.8%) were noticed only for F₆ and F₇. Except δ -guaïene (5.0%), the major compounds of F₆ volatile extract were qualitatively identical to those identified by Alitonouet *al.* in 2004 [15] in the *L. camara* leaves from Abomey-calavi. The work realized by Adelekeet *al.* on the essential oil extracted from the flowers and leaves of *L. camara* collected in Nigeria in 2004 brought back the sabinene (19.6-21.5%), 1,8-cineole (12.6-14.8%), β -caryophyllene (12.7-13.4%), α -humulene (5.8-6.3%) as major odoriferous constituents also identified in fractions F₆, F₇, F₈ [27]. In contrast, the essential oils obtained from the *L. camara* leaves collected in Cuba and China, the main compounds highlighted were respectively the (E)-nerolidol (43.4%) [28] and germacrene-D [29]. For cons, the chemical compositions of essential oils studied differ from that of the volatile extract of the flowers of Madagascar marked by the presence of davanon (22.6-25.9%) [16].

CONCLUSION

The results obtained at the end of this work deal with the chemical compositions of essential oils from *P.guajava* leaves and those of *L. camara* collected in several localities in the south of Benin and their physicochemical characteristics. The chemical profiles observed did not remain constant between two samples of the same botanical variety and by an essential oil to another. The *P. guajava* essential oils are especially rich in varied sesquiterpenoids compounds whereas those of *L. camara* are marked at the same time by the presence of hydrogenated and oxygenated monoterpenes. The rotatory powers values calculated show that the essential oil samples belonging to the same botanical species may have the

same behavior towards the polarized light. These values are on average identical for samples of *L. camara* essential oil analyzed. As for the refractive index, the results seem to show homogeneity within the same botanical species. At the level of the density, the values are not homogeneous. They could be in correlation with the origin of each essential oil. Subsequent work extended to other sites of harvested of *P. guajava* and *L. camara* leaves will allow to verify this hypothesis.

REFERENCES

- [1] Pino JA, Agüero J, Marbot R., Fuentes V. *J Essent Oil Res* 2001; 13(1): 61-62.
- [2] Ogunwande IA, Olawore NO, Adeleke KA, Ekundayo O and Koenig WA. *FlavFragr J* 2003; 18: 136-138.
- [3] da Silva JD, Luz AIR, da Silva MHL, Andrade EHA, Zoghbi MDGB and Maia JGS. *FlavFragr J* 2003; 18: 240-243.
- [4] Chen HC, Sheu MJ, Lin LY and Wu CM. *J Essent Oil Res* 2007; 19: 345-347.
- [5] Adjanohoun EJ. *Médecine traditionnelle et Pharmacopée, Contribution aux études ethnobotaniques et floristiques en République Populaire du Bénin*, ACCT, Paris, 1989.
- [6] *Flore Analytique du Benin*, A Akoegninou, WJ Van Der Burg, LLG Van Der Maessen (Eds.), Backhuys Publishers, 2006, p 1034.
- [7] Soares FD, Pereira T, Marques MOM, Monteiro AR. *Food chemistry* 2007; 100: 15-21.
- [8] Oliver-Bever M. In *Medicinal Plants in Tropical West Africa*. Cambridge University Press: Cambridge 1986; 134.
- [9] Xiao-duo J, Quan-Long P, Garrafo HM, and Pannell LK. *J Essent Oil Res* 1991; 3: 187-189.
- [10] Adam F, Vahirua-Lechat I, Deslandes E, Menut C. *J Essent Oil Res* 2011; 23: 98-101.
- [11] Petard P, *Plantes utiles de Polynésie*, Ed. revue augmentée et illustrée 1986; 244 : Ed. Haere Po No Tahiti, Papeete, French Polynesia.
- [12] Sagraero-Nieves L, Bartley JP, Provis-Schwede A. *FlavFragr J* 1994; 9(3): 135-137.
- [13] Ngassoum MB, Yonkeu S, Jirovetz L, Buchbauer G, Schmaus G and Hammerschmidt FJ. *FlavFragr J* 1999; 14: 245-250.
- [14] Kobaisy M, Tellez MR, Webber CL, Dayan FE, Schrader KK and Wedge DE. *J Agric Food Chem* 2001; 49: 3768-3771.
- [15] Alitonou G, Avlessi F, Bokossa I, Ahoussi E, Dangou J, Sohounhloúé DCK. *CR Chimie* 2004; 7: 1101-1105.
- [16] Randrianalijaona JA, Ramanoelina AR, Rasoarahona JRE, Gaydou EM. *Analyticachimica Acta* 2005; 545: 46-52.
- [17] Chowdhury JU, Nandi NC and Bhuiyan MD. *Bangladesh J Bot* 2007; 36(2): 193-194.
- [18] Sousa EO, Colares AV, Rodrigues FFG, Campos AR, Lima SG and Costa JGM. *Rec Nat Prod* 2010; 4(1): 31-37.
- [19] Rabindra KS, Balendra T. *International Journal of Pharmaceutical Research and Development* 2011; 3(7): 51-55.
- [20] *British Pharmacopoeia*, 11. P. A. HMSO: London, 1980.
- [21] Rosch P, Popp J, Kiefer W. *J Mol Struct* 1999; 121: 480-481.
- [22] Adams RP, *Identification of essential oils by ion trap mass spectrometry*. Academy Press, Inc, New-York, 1989.



- [23] Swigar AA, Silverstein RM. Monoterpenes, Infrared, Mass, NMR Spectra and Kovats Indices, Aldrich Chem. Co. Milwaukee, WI, USA. 1981.
- [24] Sonibare OO and Effiong I. African J Biotechnology 2008; 7: 2618-2620.
- [25] Ouamba JM, Ouabonzi A, Ekouya A, Bessiere JM, Menut C, Abena AA and Banzouzi JT. FlavFragr J 2006; 21: 158-161.
- [26] Hernández T, Canales M, Avila JG, García AM, Martínez A, Caballero J, Romo de Vivar A and Lira R. J Ethnopharmacol 2005; 96: 551-554.
- [27] Adeleke AK, Ekundayo O, Paul C, Koenig WA, Eshilokun AO, Yadua P. J Essent Oil Res 2004; 16: 582-584.
- [28] Pino JA, Marbot R, Rosado A, Romeu C, Pilar Marti M. J Essent Oil Res 2004; 18: 216-218.
- [29] Sundufu AJ and Shoushan H. FlavFragr J 2004; 19: 229-232.